# INHIBITION OF PEPTIDOGLYCAN TRANSPEPTIDASE BY BETA-LACTAM ANTIBIOTICS: STRUCTURE-ACTIVITY RELATIONSHIPS

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The inhibitory activities of representative  $\beta$ -lactam compounds, such as penicillins G and N, cephalosporins C and G, clavulanic acid, nocardicin A and thienamycin against *Escherichia coli* KN-126 and *Bacillus megaterium* KM peptidoglycan transpeptidases were studied. Their modes of action against *E. coli* are discussed on the basis of the results and the published binding data for penicillin binding proteins. The effects of modifications at position 3 and 7 of the cephalosporin and those at  $\alpha$ -carbon of the benzyl side-chain of cephalosporin G and penicillin G were studied. The introduction of an amino group at this position in cephalosporin G together with the removal of an acetoxy group from the acetoxymethyl group at position 3 reduced the inhibitory activity against *E. coli* transpeptidase considerably. The activity was restored by the replacement of the methyl group at position 3 of cephalexin with chlorine. The restoration was accompanied by about 15-fold increase in the lytic activity of cephachlor against *E. coli*.

Peptidoglycan transpeptidase (TPase) catalyzes the formation of cross-links at the final stage of bacterial cell wall synthesis<sup>1,2,3)</sup>. It is one of the possible lethal targets of beta-lactam antibiotics in *Escherichia coli*<sup>1,2,4)</sup>.  $\beta$ -Lactam antibiotics exert their physiological effects by binding to membrane proteins involved in morphogenesis<sup>5~7)</sup>. At least six penicillin binding proteins (PBPs) were found in the inner membrane of *E. coli* K-12. Among them the following PBPs are thought to be essential for growth<sup>6,8,9)</sup>. PBPs 1A and 1Bs are involved in cell elongation and are interchangeable in functions. Their joint inhibition by  $\beta$ -lactams results in cell lysis and spheroplast formation. PBPs 2 and 3 are involved in the maintenance of shape and in the cell division process, respectively. The TPase detectable in the membrane fraction from *E. coli* was shown to correspond to PBP 1Bs<sup>4,9,10)</sup>.

Since the demonstration of the inhibition of *E. coli* TPase by  $\beta$ -lactam antibiotics<sup>1,2)</sup> many workers have reported the inhibition in different strains of *E. coli* and in other species of bacteria. However, most of these studies are either not extensive or are unreliable because of the assay methods used for the determinations of cross-linking<sup>10,11)</sup>. We have studied the effect of many natural and semi-synthetic  $\beta$ -lactam antibiotics on the *in vitro* TPase in *E. coli* KN-126 and *Bacillus megaterium* KM with our improved method<sup>10,12)</sup>. The results will be reported here and discussed in relation to the structureactivity relationships.

### **Methods and Materials**

Organisms, growth and preparation of particulate fractions

*E. coli* KN-126 and *B. megaterium* were grown, harvested and stored as described previously<sup>10,12</sup>. Particulate fractions were prepared from the frozen cells by alumina grinding as previously reported<sup>10</sup>.

Assay of polymerization and cross-linking

The standard reaction mixture contained 0.125 M tris(hydroxymethyl) aminomethane-hydrochloride

(pH 8.5),  $0.0125 \text{ M } \text{MgCl}_2$ ,  $2.8 \times 10^{-5} \text{ M } [^{14}\text{C}]$ -UDP-N-acetylglucosamine (150  $\mu$ Ci/ $\mu$ mol),  $1.5 \times 10^{-4} \text{ M}$ UDP-N-acetylmuramylpentapeptide (prepared as described<sup>10</sup>), 16.7% (vol/vol) glycerol, and particulate fractions (*ca* 170  $\mu$ g as protein for *E. coli* and 30~60  $\mu$ g for *B. megaterium*). The mixture was incubated at 25°C for 15 or 30 minutes with *E. coli* particulate fraction and for 10 minutes with *B. megaterium* particulate fraction. The assay of polymerization and cross-linking were done as described<sup>10</sup>). The degree of cross-linking is expressed as the percentage of cross-linked disaccharide-peptide(s).

## Materials

The following antibiotics were used: penicillin G (Meiji Seika, Tokyo), penicillin N (Division Labs., Michigan Department of Health, Lansing), cephalosporin C, A-16884, A-16886 I, A-16886 II, 7-mandelo aminocephalosporanic acid, 3 N-methyltetrazolyl cephalosporin G, cephachlor, cefamandole (Lilly Research Labs., Indianapolis), nocardicin (Fujisawa Pharmaceuticals Co., Osaka), clavulanic acid, 87/359 (Glaxo Group Laboratories Ltd., Greenford), thienamycin, cefoxitin, cephamycin C (Merck Sharp & Dohme Research Labs., Rahway), cephalosporin G (kindly prepared by Dr. C. M. CIMARUSTI of E. R. Squibb & Sons (Princeton, New Jersey) and by Mr. A. SATO of this institution). UDP-N-acetyl-D-[U-<sup>14</sup>C]glucosamine (300 mCi/mmol) was purchased from Radiochemical Centre (Amersham, England).

### **Results and Discussion**

## Effects of Representative $\beta$ -Lactam Compounds

Whereas all  $\beta$ -lactam compounds studied except nocardicin are moderate to very strong inhibitors of *B. megaterium* transpeptidase, only penicillin G (Pen G) and thienamycin are strong inhibitors of *E. coli* TPase. Clavulanic acid was a poor inhibitor of *E. coli* TPase and strong inhibitor of *B. megaterium* TPase. The hydrophilic  $\alpha$ -aminoadipyl side-chain reduced considerably the inhibitory activity of both penicillin N and cephalosporin C toward *B. megaterium* TPase. It had little effect on the inhibitory activity of the cephalosporin nucleus to *E. coli* TPase (Table 1).

The above results together with published data for the penicillin binding proteins in *E. coli* KN-126<sup>7,13)</sup> indicate that the action of the representative  $\beta$ -lactam compounds can be characterized as follows. Pen G has a nearly equal affinity toward the three possible lethal targets in *E. coli*, namely PBP 1Bs, PBP 2 and PBP 3. Cephalosporin G (CP-G) has the highest affinity toward PBP 3. Its high affinity toward PBP 1 (later resolved into 1A and 1Bs<sup>14)</sup>) can be explained by the affinity toward PBP 1A, which is functionally interchangeable with 1Bs but has higher affinity toward  $\beta$ -lactams. The affinity toward PBP 2 is the least, (This situation will be described as follows: 3>1Bs>2). Like mecillinam,

β-Lactam	Concentrations required to inhibit cross-linking 50 % (µg/ml)		
-	E. coli	B. megaterium	
Penicillin G	4.0	0.052	
N	48.0	0.7	
Cephalosporin G	60	0.33	
С	60	16	
Clavulanic acid	460	0.76	
Nocardicin A	80	125	
Thienamycin	2.4	0.024	

Table	Eff	fects	of	represe	entative	β-lact	tam	com-
pounds	on	pep	tide	glycan	cross-lin	nking	in	vitro.

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clavulanic acid is a specific inhibitor of PBP 2 function(s)<sup>13)</sup> and its affinities toward other PBPs will not contribute to its antimicrobial action against *E. coli* (2  $\gg$  1Bs, 3). Thienamycin has the highest affinity toward PBP 2, but its high inhibitory activity against the *in vitro* TPase (1Bs) may contribute to its antimicrobial activity (2>1Bs $\gg$ 3).

# Effects of the Introduction of a $7-\alpha$ -Methoxy Group

The introduction of a 7-methoxy group into cephalosporins increased the inhibitory

activity toward E. coli TPase as reported previously<sup>15)</sup> (Table 2). The introduction did not affect the activity toward B. megaterium TPase. Since an increased inhibitory activity against E. coli TPase was not always reflected in the antimicrobial activity, a poor permeability was assumed<sup>15)</sup>. Recently CURTIS et al. have studied the affinities and antibacterial activities of several 7-methoxycephalosporins in E. coli K-12 (DCO) and its permeability mutant DC2 and have concluded that the decrease in the antibacterial activity is due to a reduced affinity for an essential PBP and not to less favourable permeability properties<sup>16)</sup>. Thus the situation seems to be complex. In some cases the introduction of a 7-methoxy group increases the affinity toward

β-Lactam	Concentrations required to inhibit cross-linking 50 % (µg/ml)			
	E. coli	B. megaterium		
Cephalosporin C	60	16		
A-16884	8	17		
A-16886 II	30	2		
I	8	1.5		
87/359	12.5	_		
Cefoxitin	4.1	0.15		

Table 2. Effects of introduction of a 7- $\alpha$ -methoxy group into cephalosporins.

A-16886 I=cephamycin C (7-(5-amino-5-carboxy-valeramido)-7-methoxy-3-carbamoyloxymethyl-3-cephem-4-carboxylic acid).

Each pair differs only by the presence of a 7-methoxy group.

-, not determined.

PBP 1Bs (the *in vitro* TPase) with a considerable decrease in the affinity toward PBP 3. In other cases, the introduction decreases the affinities toward both PBPs, but the decrease in the affinity toward PBP 1Bs is less<sup>16)</sup>. In rare cases it increases the affinity toward PBP 1Bs with little or no decrease in the affinity toward PBP 3. Whichever the case, the ratio of the affinity toward PBP 1Bs to that of PBP 3 will be increased by the introduction of a methoxy group to cephalosporins. Consequently the lower concentration range where elongation is mainly observed with corresponding 7-H cephalosporins becomes narrower or disappears and lysis is observed as the earliest event.

# Effects of Substitutions at Position 3 of Cephalosporins

Cephalosporin C (CP-C), deacetyl CP-C and deacetoxy CP-C show decreasing inhibitory activity against *E. coli* TPase in this order (Table 3). The substitution of the acetyl group with a carbamoyl group increases the inhibitory activity slightly against *E. coli* TPase and significantly against *B. megaterium* TPase. This relationship will also hold true for the substitution at position 3 of cephamycins

β-Lactam	Functional group at position 3	Concentrations required to inhibit cross-linking 50 % (µg/ml)		
		E. coli	B. megaterium	
Cephalosporin C	acetoxymethyl	60	16	
Deacetyl CP-C	hydroxymethyl	360	12.5	
Deacetoxy CP-C	methyl	720	20	
A-16886 II	carbamoyloxymethyl	30	2	
Cephaloglycin	acetoxymethyl	167	0.5	
Cephalexin	methyl	500	1	
Cephachlor	chloro	12	0.09	
Cephalosporin G	acetoxymethyl	60	0.33	
CP-G: 3-tetrazolyl thiomethyl	N-methyltetrazolylthiomethyl	3.9	-	

Table 3. Effects of substitutions at C-3 position of cephalosporins.

CP-G: 3-tetrazolylthiomethyl, 7-phenylacetoamido-3-N-methyltetrazolylthiomethyl-3-cephem-4-carboxylic acid. —, not determined. (data not shown). The substitution of the acetoxymethyl group with chlorine (cephachlor) or a N-methyl tetrazolyl-thiomethyl group considerably increased the inhibitory activity against *E. coli* TPase.

# Effects of Substitutions at the $\alpha$ -Carbon of the

Benzyl Side-Chain in CP-G and Pen G

The introduction of an  $\alpha$ -amino group to the benzyl side chain of both a penicillin and a cephalosporin resulted in a reduction of the inhibitory activity against both *E. coli* and *B. megaterium* TPases (Table 4). Also, the introduction of a carboxyl or sulfonyl group into Pen G at this position reduced the inhibitory activity against TPases from both organisms. The introduction of a hydroxy group into the corresponding position of CP-G slightly decreased the inhibitory activity against *E. coli* TPase. Thus Fig. 1. Effect of cephalexin (CEX) and cephachlor (CCL) on growth of *E. coli* KN-126.

Cells were grown at 37°C in a bouillon medium containing 2% dehydrated nutrient broth (Kyokuto Pharmaceuticals Co., Tokyo) and 0.5% yeast extract (pH 7). Numbers show the amount of antibiotics ( $\mu$ g/ml) added at times indicated by the arrow.



the high inhibitory activity of cephamandole against *E. coli* TPase<sup>10</sup> can be solely explained by the presence of a N-methyltetrazolylthiomethyl group at position 3. The introduction of an  $\alpha$ -amino group into the benzyl side-chain of Pen G to give ampicillin did not affect or only slightly improved the affinity toward PBP 3, which is the most sensitive target toward ampicillin<sup>7,17</sup>. Thus by this criterion the intrinsic activity of ampicillin against *E. coli* is more or less similar to that of Pen G, though their minimum inhibitory concentrations (MIC) differ widely.

# Comparison of Cephachlor with Cephalexin

The introduction of an  $\alpha$ -amino group into the benzyl side-chain of CP-G together with the removal of an acetoxy group from the acetoxymethyl group at position 3 of CP-G considerably reduced the inhibitory activity of the resultant compound (cephalexin) against *E. coli* TPase (Table 3). These modifications did not impair the affinity toward PBP 3 and might increase it. Thus cephalexin remains

β-Lactam	Substitution at benzylic carbon	Concentrations required to inhibit cross-linking 50 % (µg/ml)		
		E. coli	B. megaterium	
Penicillin G		4.0	0.052	
Ampicillin	$\rm NH_2$	10	0.7	
Carbenicillin	СООН	23.2	0.42	
Sulbenicillin	SO <sub>3</sub> H	29	0.32	
Cephalosporin G		60	0.33	
Cephaloglycin	$\rm NH_2$	167	0.5	
Mandelo 7-ACA	ОН	96		

Table 4. Effects of substitutions at the  $\alpha$ -carbon of the benzyl side-chain.

Mandelo-7-ACA, 7-phenylhydroxyacetoamido-3-acetoxymethyl-3-cephem-4-carboxylic acid.

a potent antimicrobial agent against *E. coli*. The substitution of the methyl group at position 3 of cephalexin with chlorine increased the inhibitory activity against *E. coli* TPase fortyfold and *B. mega-terium* TPase tenfold. The affinities toward PBP 1Bs and 3 were increased 55-fold and 4-fold respectively<sup>17)</sup>. With these changes, the antibacterial activity increased only twofold or so in terms of MIC<sup>17)</sup>. However, cells of growing *E. coli* KN-126 lysed rapidly at 25  $\mu$ g of cephachlor or less per ml, whereas they lysed at 250  $\mu$ g of cephalexin per ml (Fig. 1). ROLINSON<sup>18)</sup> observed lysis of filamentous cells of *E. coli* at low concentrations of cephalexin (2~50×MIC) and rapid lysis of short cells at higher concentrations (around 200×MIC). He suggested that the lysis of these two forms of *E. coli* might be due to two different processes. Indeed, the rapid lysis can be explained by the inhibition of the *in vitro* TPase (PBP 1Bs), while the lysis at low concentrations will be ascribed to the inhibition of PBP 3 function(s) (septum formation). How the improvement in the inhibitory activity of cephachlor against *E. coli* TPase is reflected in the clinical situation remains to be seen.

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### References

- ARAKI, Y.; R. SHIRAI, A. SHIMADA, N. ISHIMOTO & E. ITO: Enzymatic synthesis of cell wall mucopeptide in a particulate preparation of *Escherichia coli*. Biochem. Biophys. Res. Commun. 23: 466~472, 1968
- IZAKI, K.; M. MATSUHASHI & J. L. STROMINGER: Biosynthesis of the peptidoglycan of bacterial cell walls. XIII. Peptidoglycan transpeptidase and D-alanine carboxypeptidase. J. Biol. Chem. 243: 3180~3192, 1968
- 3) WICKUS, G. G. & J. L. STROMINGER: Penicillin sensitive transpeptidation during peptidoglycan biosynthesis in cell-free preparations from *Bacillus megaterium*. J. Biol. Chem. 247: 5307~5311, 1972
- NAKAGAWA, J.; S. TAMAKI & M. MATSUHASHI: Purified penicillin binding proteins 1Bs from *Escherichia coli* membrane showing activities of both peptidoglycan polymerase and peptidoglycan crosslinking enzyme. Agr. Biol. Chem. 43: 1379~1380, 1979
- 5) SPRATT, B. G.: Distinct penicillin-binding proteins involved in the division, elongation and shape of *Escherichia coli*. Proc. Natl. Acad. Sci. U.S.A. 72: 2999~3003, 1975
- 6) SPRATT, B. G.: Penicillin-binding proteins of *Escherichia coli*: General properties and characterization of mutants. pp. 182~190. *In* D. SCHLESSINGER (*ed*), Microbiology. American Society for Microbiology, Washington, D. C., 1977
- 7) SPRATT, B. G.: Properties of the penicillin-binding proteins of *Escherichia coli* K-12. Eur. J. Biochem. 72: 341~352, 1977
- SUZUKI, H.; Y. NISHIMURA & Y. HIROTA: On the process of cellular division in *Escherichia coli*: A series of mutants of *E. coli* altered in the penicillin binding proteins. Proc. Natl. Acad. Sci. U.S.A. 75: 664~668, 1978
- 9) TAMAKI, S.; T. NAKAJIMA & M. MATSUHASHI: Thermosensitive mutation in *Escherichia coli* simultaneously causing defects in penicillin-binding protein 1Bs and in enzyme activity for peptidoglycan synthesis *in vitro*. Proc. Natl. Acad. Sci. U.S.A. 74: 5472~5476, 1977
- ΟΚΑ, Τ. & H. FUJITA: Effect of β-lactam antibiotics on *in vitro* peptidoglycan cross-linking by a particulate fraction from *Escherichia coli* K-12 and *Bacillus megaterium*. Antimicr. Agents & Chemother. 14: 625~ 627, 1978
- 11) OKA, T.: The mechanism of action of  $\beta$ -lactam antibiotics. (in Japanese) Kagaku to Seibutsu 17: 625~633, 1979
- ΟκΑ, Τ.: Mode of action of penicillins in vivo and in vitro in Bacillus megaterium. Antimicr. Agents & Chemother. 10: 579~591, 1976
- 13) SPRATT, B. G.; V. JOBANPUTRA & W. ZIMMERMANN: Binding of thienamycin and clavulanic acid to the

penicillin-binding proteins of *Escherichia coli* K-12. Antimicr. Agents & Chemother. 12: 406~409, 1977
14) SPRATT, B. G.; V. JOBANPUTRA & U. SCHWART. Mutants of *Escherichia coli* which lack a component of 1 are viable. FEBS Lett. 79: 660~663, 1976

- 15) Ho, P. P. K.; R. D. TOWNER, J. M. INDELICATO, W. J. WILHAM, W. A. SPITZER & G. A. KOPPEL: Biochemical and microbiological studies on 7-methoxycephalosporins. J. Antibiotics 26: 313~314, 1973
- 16) CURTIS, N. A. C.; G. W. Ross & M. BOULTON: Effect of 7-α methoxy substitution of cephalosporins upon their affinity for the penicillin-binding proteins of *E. coli* K12: Comparison with antibacterial activity and inhibition of membrane bound model transpeptidase activity. J. Antimicrob. Chemother. 5: 391~398, 1979
- 17) CURTIS, N. A. C.; D. ORR, G. W. Ross & M. G. BOULTON: Affinities of penicillin-binding proteins of Escherichia coli K-12 and their antibacterial activity. Antimicr. Agents & Chemother. 16: 533~539, 1979
- ROLINSON, G. N.; A. C. MACDONALD & D. A. WILSON: Bactericidal action of β-lactam antibiotics on *Escherichia coli* with particular reference to ampicillin and amoxycillin. J. Antimicrob. Chemother. 3: 541 ~ 553, 1977